Slide preparation for interphase FISH

1. We recommend pre-cleaning microscope slides (even if advertised as “pre-cleaned” with a 2’ rinse in 100% ethanol
2. Allow the slides to dry completely via air-drying
3. Incubate slides in a 0.01% (v/v) poly-L-Lysine solution for 5’ in a plastic coplin jar
4. Allow slides to completely dry via air-drying
5. Prepare a cell suspension of 0.5-2 x 10^6 cells/ml in growth media
6. Add 100 µl of cell suspension per slide roughly in the center
7. Allow the cells to adhere for 1-3 hours at the growth temperature of the cell line (e.g. 37°C, 5% CO_2 for mammalian cells; 25°C atmospheric CO_2 for insect cells)
8. (*All steps now at room temp*) Rinse slides briefly in 1X PBS
9. Fix slides for 5-15’ in 4% (v/v) paraformaldehyde in 1X PBS
10. Rinse briefly in 1X PBS in a coplin jar
11. Incubate for 5’ in 2X SSCT in a coplin jar
12. Incubate for 5’ in 2X SSCT + 50% (v/v) formamide in a coplin jar
13. Transfer to a fresh coplin jar containing 2X SSCT + 50% formamide for storage at 4°C.
14. Slides are typically best used within 1-2 weeks of creation.

2X SSCT - 0.3 M NaCl, 0.03 M NaCitrate, 0.1% Tween-20